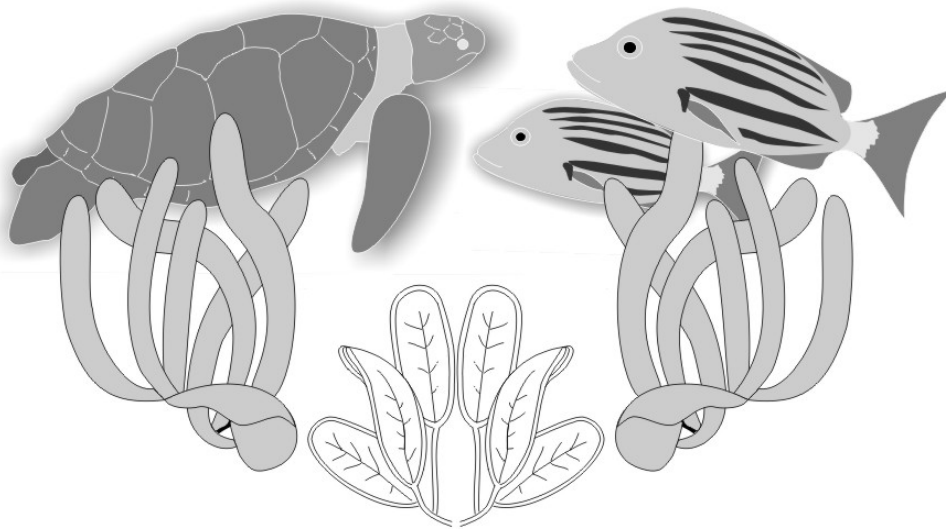
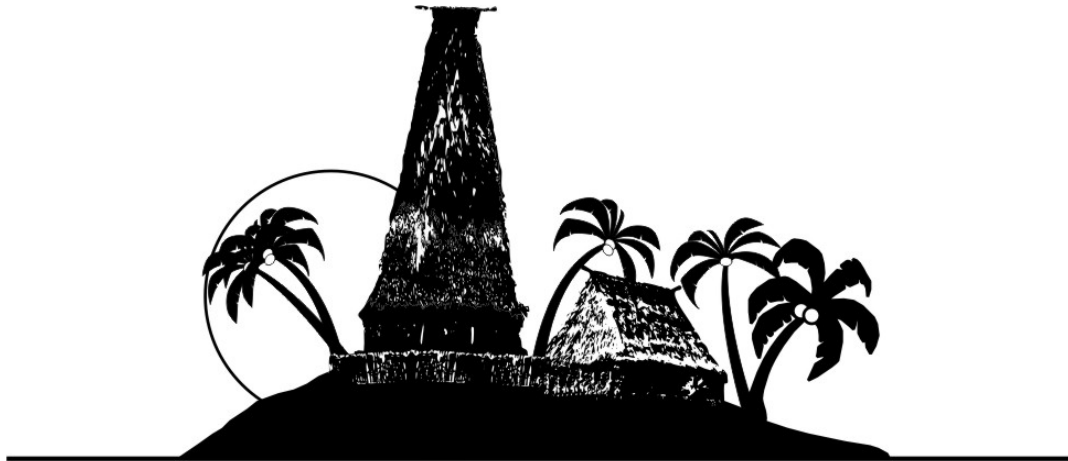


Seagrass-Watch

Guidelines for Monitoring Seagrass Habitats in the
Fiji Islands



Corpus Christi Teachers College, Laucala Bay, Suva

16th June 2007

Len McKenzie & Rudi Yoshida



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Overview

Seagrass-Watch is the largest scientific, non-destructive, seagrass assessment and monitoring program in the world.

Often governments are unable to protect and conserve seagrass meadows without the assistance of local communities (e.g., local residents, schools, tertiary institutions, non-government organisations). Seagrass-Watch is a monitoring program that brings citizens and governments together for seagrass conservation. It identifies areas important for seagrass species diversity and conservation. The information collected can be used to assist the management of coastal environments and to prevent significant areas and species being lost.

Monitoring seagrass resources is important for two reasons: it is a valuable tool for improving management practices; and it allows us to know whether resource status and condition is stable, improving or declining. Successful management of coastal environments (including seagrass resources) requires regular monitoring of the status and condition of natural resources.

Early detection of change allows coastal management agencies to adjust their management practices and/or take remedial action sooner for more successful results. Monitoring is important in improving our understanding of seagrass resources and to coastal management agencies for:

- *Exposing coastal environmental problems before they become intractable,*
- *Developing benchmarks against which performance and effectiveness can be measured,*
- *Identifying and prioritising future requirements and initiatives,*
- *Determining the effectiveness of management practices being applied,*
- *Maintaining consistent records so that comparisons can be made over time,*
- *Developing within the community a better understanding of coastal issues,*
- *Developing a better understanding of cause and effect in land/catchment management practices,*
- *Assisting education and training, and helping to develop links between local communities, schools and government agencies, and*
- *Assessing new management practices*

A rapid method for monitoring seagrass resources is used in the Seagrass-Watch program. Seagrass-Watch monitoring efforts are vital to assist with tracking global patterns in seagrass health, and assess the human impacts on seagrass meadows, which have the potential to destroy or degrade these coastal ecosystems and decrease their yield of natural resources. Responsive management based on adequate information will help to prevent any further significant areas and species being lost. To protect the valuable seagrass meadows along our coasts, everyone must work together.

The goals of the Seagrass-Watch program are:

- *To educate the wider community on the importance of seagrass resources*
- *To raise awareness of coastal management issues*
- *To build the capacity of local stakeholders in the use of standardised scientific methodologies*
- *To conduct long-term monitoring of seagrass & coastal habitat condition*
- *To provide an early warning system of coastal environment changes for management*
- *To support conservation measures which ensure the long-term resilience of seagrass ecosystems.*

As part of this workshop we will

- *learn seagrass taxonomy*
- *discuss the present knowledge of seagrass ecology,*
- *discuss the threats to seagrasses*
- *learn techniques for monitoring seagrass resources*
- *provide examples of how Seagrass-Watch assists with the management of impacts to seagrass resources and provides an understanding of their status and condition.*

The following information is provided as a training guide and a reference for future Seagrass- Watch monitoring activities. For further information, please do not hesitate to contact us at

Seagrass-Watch HQ

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Workshop leaders



Len McKenzie

Len is a Principal Scientist with the Queensland Department of Primary Industries & Fisheries and Seagrass-Watch Program Leader. He is also chief investigator for the MTSRF task on the condition, trend and risk in coastal seagrass habitats, Task Leader of the RWQPP Marine Monitoring Programme - Seagrass Monitoring and project leader for a series of projects involving the assessment and sustainable use of coastal fisheries habitat. Len has 19 years experience as a research scientist on seagrass ecology, assessment and fisheries habitats. This includes experience within Australia and overseas in seagrass research, resource mapping/ assessment and biodiversity. He has provided information on seagrass communities that has been vital in management of seagrass resources of the Great Barrier Reef and also at the state, national and international levels. He has also advised on fisheries and coastal resource-use issues for managers, fishing organisations, conservation and community groups. Len is also the Secretary of the World Seagrass Association.

Current Projects

- Seagrass-Watch
- Status and mapping of seagrass resources in Queensland
- Assessment of primary and secondary productivity of tropical seagrass ecosystems
- Investigations on the macrofauna associated with seagrass meadows
- Great Barrier Reef Water Quality Protection Plan – marine monitoring program: seagrass



Rudi Yoshida

Rudi is a Scientific Assistant with the Queensland Department of Primary Industries & Fisheries. Rudi has over 10 years experience in seagrass related research and monitoring. He is also a core member of Seagrass-Watch HQ, and ensures data submitted is managed and QA/QC protocols applied. He is also responsible for maintenance of the Seagrass-Watch website.

Current Projects

- Seagrass-Watch
- Great Barrier Reef Water Quality Protection Plan – marine monitoring program: seagrass



Agenda

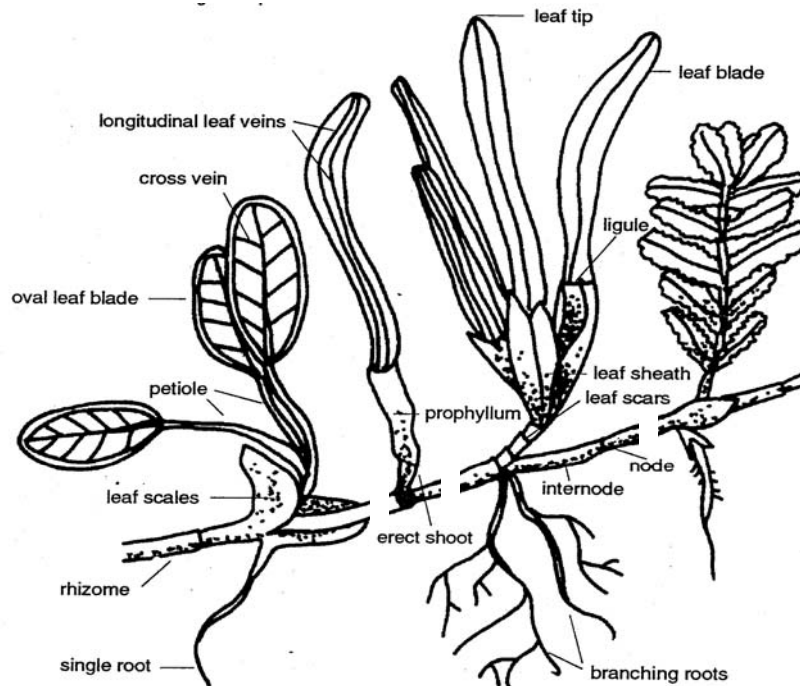
Saturday 16th June 2007

Morning	0920 – 0930	Welcome & Introduction - <i>TBA & Len</i>
	0930 – 0945	Seagrass Biology & Identification - <i>Len</i>
	0945 – 1030	Laboratory exercise: Seagrass Identification & how to prepare a seagrass press specimen - <i>Len & Rudi</i>
	1030 – 1045	<i>Morning Tea</i>
	1045 – 1125	Seagrass Ecology and Threats - <i>Len</i>
	1125– 1130	Seagrass monitoring - <i>Len</i>
	1130– 1230	Seagrass-Watch - <i>Len</i>
Afternoon	1230 - 1300	<i>Lunch</i>
	1315 - 1530	<p>Field exercise:</p> <p>Seagrass-Watch monitoring SV1 – <i>Len & Rudi</i></p> <p>Where: Nesese</p> <p>What to bring:</p> <ul style="list-style-type: none"> • <i>hat, sunscreen (Slip! Slop! Slap!)</i> • <i>dive booties or old shoes that can get wet</i> • <i>drink/refreshments</i> • <i>polaroid sunglasses (not essential)</i> • <i>enthusiasm</i> <p><i>You will be walking across a seagrass meadow exposed with the tide, through shallow water. It may be wet and muddy!</i></p> <p>Wrap up (<i>Corpus Christi Teachers College</i>)</p>
	1530 - 1600	<ul style="list-style-type: none"> • Feedback • <i>Presentation of certificates</i>

Low tide: 1:30pm 0.2m

Background

Seagrasses are specialised marine flowering plants that have adapted to the nearshore environment of most of the world's continents. Most are entirely marine although some species cannot reproduce unless emergent at low tide. Some seagrasses can survive in a range of conditions encompassing fresh water, estuarine, marine, or hypersaline. There are relatively few species globally (about 60) and these are grouped into just 13 Genera and 5 Families.



Composite illustration demonstrating morphological features used to distinguish main seagrass taxonomic groups.

from Lanyon (1986)

Various common names are applied to seagrass species, such as turtle grass, eelgrass, tape grass, spoon grass and shoal grass. These names are not consistently applied among countries. Coastal communities would almost certainly recognise the term "turtle grass" as referring to the shallow subtidal and intertidal seagrasses.

There is now a broad understanding of the range of species and seagrass habitats. Areas less well known include the southeast Pacific reefs and islands, South America, the southern Atlantic, the Indian Ocean islands, the west African coast, and Antarctica. Shallow sub-tidal and intertidal species distributions are better recorded than seagrasses in water greater than 10 m below MSL. Surveying deeper water seagrass is time consuming and expensive and it is likely that areas of deepwater seagrass are still to be located.

Tropical seagrasses occupy a variety of coastal habitats. Tropical seagrass meadows typically occur in most shallow, sheltered soft-bottomed marine coastlines and estuaries. These meadows may be monospecific or may consist of multispecies communities, sometimes with up to 12 species present within one location. The stresses and limitations to seagrasses in the tropics are generally different than in temperate or subarctic regions. Temperature related impacts most often result from high water temperatures or overexposure to warm air; osmotic impacts result from hypersalinity due to evaporation; radiation impacts result from high irradiance and UV exposure.



The depth range of seagrass is most likely to be controlled at its deepest edge by the availability of light for photosynthesis. Exposure at low tide, wave action and associated turbidity and low salinity from fresh water inflow determines seagrass species survival at the shallow edge. Seagrasses survive in the intertidal zone especially in sites sheltered from wave action or where there is entrapment of water at low tide, (e.g., reef platforms and tide pools), protecting the seagrasses from exposure (to heat, drying or freezing) at low tide.

Tropical seagrasses are important in their interactions with mangroves and coral reefs. All these systems exert a stabilizing effect on the environment, resulting in important physical and biological support for the other communities (Amesbury and Francis 1988).

Barrier reefs protect coastlines, and the lagoon formed between the reef and the mainland is protected from waves, allowing mangrove and seagrass communities to develop. Seagrasses trap sediment and slow water movement, causing suspended sediment to fall out. This trapping of sediment benefits coral by reducing sediment loads in the water.

Mangroves trap sediment from the land, reducing the chance of seagrasses and corals being smothered. Sediment banks accumulated by seagrasses may eventually form substrate that can be colonized by mangroves. All three communities trap and hold nutrients from being dispersed and lost into the surrounding oceanic waters.

The habitat complexity within seagrass meadows enhances the diversity and abundance of animals. Seagrasses on reef flats and near estuaries are also nutrient sinks, buffering or filtering nutrient and chemical inputs to the marine environment. The high primary production rates of seagrasses are closely linked to the high production rates of associated fisheries. These plants support numerous herbivore- and detritivore-based food chains, and are considered as very productive pastures of the sea. The associated economic values of seagrass meadows are very large, although not always easy to quantify.

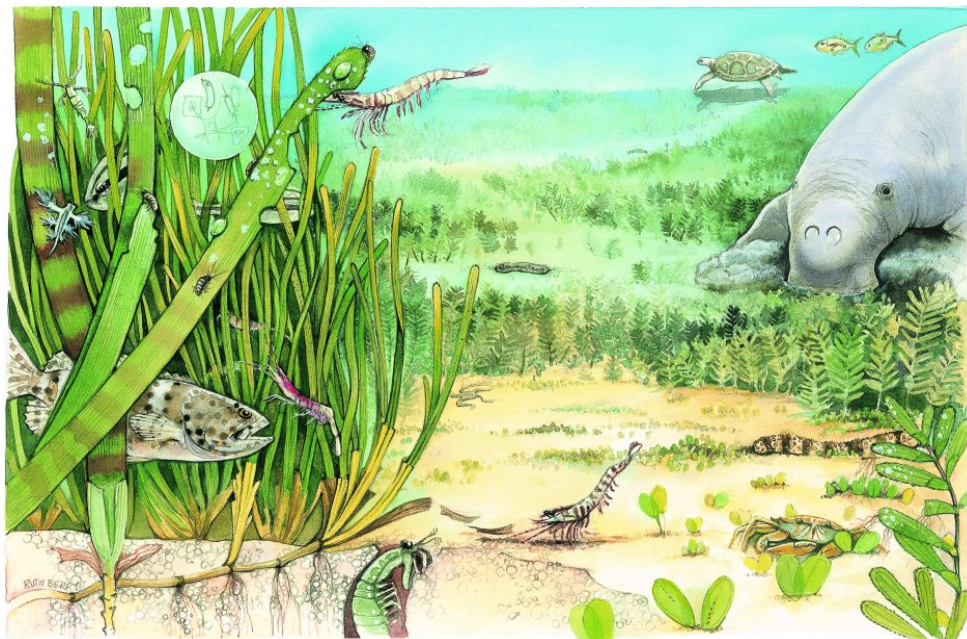
Tropical seagrass meadows vary seasonally and between years, and the potential for widespread seagrass loss has been well documented (Short and Wyllie-Echeverria 1996). The causes of loss can be natural such as cyclones and floods, or due to human influences such as dredging, agricultural runoff, industrial runoff or oil spills.

Loss of seagrasses has been reported from most parts of the world, sometimes from natural causes, e.g., high energy storms, or "wasting disease". More commonly, loss has resulted from human activities, e.g., as a consequence of eutrophication or land reclamation and changes in land use. Anthropogenic impacts on seagrass meadows are continuing to destroy or degrade these coastal ecosystems and decrease their yield of natural resources.

It is important to document seagrass species diversity and distribution and identify areas requiring conservation measures before significant areas and species are lost. Determining the extent of seagrass areas and the ecosystem values of seagrasses is now possible on a local scale for use by coastal zone managers to aid planning and development decisions. Knowledge of regional and global seagrass distributions are still too limited and general for broad scale protection and management. Such information is needed to minimize future impacts on seagrass habitat worldwide. With global electronic communication it is now possible to begin the process of assembling both formally published and unpublished notes on the distribution of the world's seagrasses with the eventual aim of providing a global "report card" on the distribution and status of seagrass.

With well-recorded events of seagrass loss from many coastal environments it is important to map and record the distribution of not only the location of existing seagrass but also areas of potential seagrass habitat. Such areas are generally shallow, sheltered coastal waters with suitable bottom type and other environmental conditions for seagrass growth. Potential habitat may include areas where seagrass was known to grow at some time in the past but from which it has recently been eliminated.

Spatial and temporal changes in seagrass abundance and species composition must be measured and interpreted with respect to prevailing environmental conditions. These may need to be measured seasonally, monthly, or weekly, depending on the nature of their variability, and the aims of the study. Physical parameters important to seagrass growth and survival include light (turbidity, depth), sediment type and chemistry, and nutrient levels. Detailed studies of changes in community structure of seagrass communities are essential to understand the role of these communities and the effects of disturbance on their composition, structure and rate of recovery.



[illegible]

Seagrasses of the Fiji Islands

Seagrass meadows (*veivutia*) are found intertidal and in the shallow subtidal waters of protected and soft shores throughout Fiji. They play an important role in maintaining coastal water quality and are vital in supporting coastal marine communities and maintaining diverse flora and fauna.

Seagrasses have high biological productivity, are efficient recyclers of nutrients and support a large biomass of consumers, especially those of fisheries importance. It has been suggested that 400 square metres of seagrass (10 metres long and 40 metres wide) can support 2000 tonnes of fish a year. Fiji's coastal fisheries productivity depends greatly on seagrass habitats. For example, juvenile Emperors (*kawago* - *Lethrinus nebulosus*; *sabutu*, *cabutu* - *L. atkinsoni*; *kabatia*, *kabatiko* - *L. harak*) live in the shallow, inshore areas such as seagrass and mangrove before they move to deeper water as adults (Richards *et al.* 1994). The ark shell (*kaikoso*, *qege* - *Anadara cornea*) although patchily distributed, are common in seagrass meadows of Laucala Bay (Butler 1983). It has been suggested that *kaikoso* recruits into sand/seagrass areas, though it can live in mud where the sediment is dynamic (due to floods and other causes) when older (Butler 1983).

Seagrass is also a nursery habitat for the witch prawn (*uranicakau* - *P. canaliculatus*). Post-larvae settle into seagrass meadows on the intertidal mudflats in June and November of each year, after adults spawn in the deep channels of Laucala Bay (Choy 1982). The post-larvae grow in the nursery grounds for approximately 5 months, until as juveniles they move offshore into the adult stocks, eventually mating and spawning in October-November (Choy 1982). Seagrass are also important habitats for *moci* (mangrove prawns/shrimps – *Palaemon* sp.) and octopus (*kuita*, *sulua* - *Octopus* sp.) (Lewis 1985; Richards *et al.* 1994). Seagrass is also one of the food items of rock lobsters (e.g., *uraukula*, *uraudina*, *urautamata* - *Panulirus* spp) (Lewis 1985).

Fiji's extensive pastures of seagrass are also a significant resource for green turtles in the central south Pacific region. Green turtles spend most of their adult life foraging in Fijian waters, occupying home ranges averaging 27 km², taking only brief migrations (up to 1066km) to French Polynesia, American Samoa, Tonga and Cook Islands to nest (Craig *et al.* 2004). The seagrass foraging areas in Fiji may well be providing foraging habitat for over half of the adult greens in the central South Pacific. This is possibly a consequence of lower availability of turtle food east of Fiji where most islands are small, steep and have limited areas suitable for seagrass. The need to protect such foraging areas is becoming widely recognized as a critical part of sea turtle conservation.

Five seagrass species and one subspecies are reported from the Fiji Islands: *Halodule pinifolia*, *Halodule uninervis*, *Halophila ovalis*, *Halophila ovalis* ssp. *bullosa*, *Halophila decipiens* and *Syringodium isoetifolium* (Spalding *et al.* 2003; Skelton and South 2006). Records of *Thalassia hemprichii*, *Cymodocea serrulata*, *Halophila minor*, and *Halophila ovata* credited to Fiji are either erroneous or remain to be verified. All the species within Fiji waters have an Indo-Pacific distribution, except *H. ovalis* ssp. *bullosa* which is endemic to Fiji, Tonga, and Samoa (McMillan and Bridges 1982).

Halodule pinifolia is generally found in the high intertidal to upper subtidal areas of sheltered bay, reef platforms and in high energy locations. *H. pinifolia* often forms homogenous patches or occasionally intermixes with other seagrass species including the closely related *H. uninervis*. They are easily distinguishable in the field by their much narrower blade size compared with that of *H. uninervis* (1 mm versus 4 mm).



Waycott *et al.* (2004) suggested that *H. pinifolia* and *H. uninervis* are conspecific, recognising that the plasticity of blade size is attributed to local conditions. Nevertheless, in Fiji we retain them as separate entities, as there is no sufficient evidence from Fijian material to support this merger. Future studies, both ecological and molecular, would help to clarify this.

Halodule uninervis is found from intertidal to 30m in sheltered or exposed coral reefs, in creeks and mangroves. *H. uninervis* often forms dense meadows at some sites, or is patchy and intermixed with other seagrass species (viz. *H. uninervis*, *S. isoetifolium*, or *Halophila* spp.).

Halophila ovalis is the most eurythermic of all seagrasses in Fiji and extends from the intertidal to 10-12m deep. First reported in Fiji in 1874 from Viti Levu, there are both bullate (blister or pucker-like) and smooth leaf forms (den Hartog 1970). The bullate form is recognized as a subspecies *H. ovalis* ssp. *bullosa* because it appears sufficiently distinct (McMillan and Bridges 1982). However the synonymy adopted here follows Waycott *et al.* (2004) who consider *H. ovalis* to be a complex of closely related entities whose leaves are highly plastic especially in relation to blade size, shape, colour, and texture (Waycott *et al.* 2002). The bullate forms have only been reported from Samoa, Tonga and the Fiji islands. *H. ovalis* forms dense meadows in some locations, but is frequently encountered in small patches. It tolerates a wide variety of substrata from fine muddy sand to coarse sand, mixed sandy-rubble or large boulders with sandy patches.

Halophila decipiens is a recent addition to the seagrass inventory in Fiji. It is no surprise that it occurs in Fiji, as the species is pan tropical and has been reported in New Caledonia and Tahiti from the mid-late 1800's (den Hartog 1970). Globally, *H. decipiens* is a sciophilous species which occurs from the water surface to a depth of 85m (Cargados Carajos Shoals, Mascarene Islands in the Indian Ocean). In the Fiji Islands it occurs from 10–25 m depth and has only been found growing in the fine muddy/sandy substratum along the reef channels of the Great Sea Reef (Skelton and South 2006). The plants form sparse patches and grow to 40 mm tall. *H. decipiens* is distinguished from *H. ovalis* and its subspecies *bullosa* by the presence of marginal serrations and hairs on either side of the leaf blade.

Syringodium isoetifolium is usually found in the shallow subtidal areas (1–6 m depth), with some meadows are occasionally exposed during extreme low tide on reef flats. Earliest records are from 1926 in Suva Bay (den Hartog 1970). *Syringodium* is known to be more tolerant of oxidized substrata than other seagrass species and it has been reported that *Syringodium* will take over as a pioneer after a disturbance (den Hartog, 1977). *S. isoetifolium* also has the ability to utilise a very high proportion of the available dissolved inorganic carbon compared to other seagrass species. Such an environment provides *S. isoetifolium* with a competitive advantage, especially when combined with disturbances that remove existing seagrass species.

A number of general parameters are critical to whether seagrass will occur along any stretch of coastline. These include physical parameters that regulate the physiological activity of seagrasses (temperature, salinity, waves, currents, depth, substrate and day length), natural phenomena that limit the photosynthetic activity of the plants (light, nutrients, epiphytes and diseases), and anthropogenic inputs that inhibit the access to available plant resources (nutrient and sediment loading). Various combinations of these parameters will permit, encourage or eliminate seagrass from a specific location. In Fiji, seagrasses are distributed throughout the Islands and local conditions may often determine which seagrass species are present.

VITI LEVU

Along northern Viti Levu, *H. uninervis*, *H. pinifolia*, *H. ovalis* and *S. isoetifolium* are present on intertidal mudflats and fringing reefs (e.g., Navolau village, Raki Raki) (Pers. Obs.). However, most information for Viti Levu is concentrated along the southern shores.

Coral coast

The Coral Coast stretches along an 80 km length of coast on the southern side of Viti Levu. It is the longest chain of fringing reefs in Fiji. The Coral Coast embraces Natadola Beach in the west, to Pacific Harbour further to the east. Seagrass meadows cover the nearshore areas of the fringing reef, where there is sufficient sediment depth and stability.

Seagrass meadows have been surveyed on the shallow mudflat on the north west of **Likuri** Island (Robinson Crusoe Resort), just off the start of the Coral Coast. The surveys identified *Syringodium isoetifolium* meadows which had the highest algal cover (probably *Dictyota* sp) of all the habitats examined around the island (Sykes 2003). The shallow mudflats have been extensively over-fished over a long period by village subsistence fishers, and small scale commercial fishers from the main island. It is also heavily affected by silt run-off from the nearby river. Coral health is poor. The survey was part of a project to develop a standard Environmental Impact Assessment technique that would be suitable for use at small resorts throughout the Fiji islands.

A few kilometres south, small patches of *H. uninervis* and *H. pinifolia* can be found on the fringing reefs of **Natadola**. The seagrass has very little epiphyte cover and the environment excellent water clarity. Although relatively pristine, the meadows are threatened by adjacent coastal development.

In the nearby **Cuvu** Bay, there are large seagrass meadows, but they are also threatened by turbid flood waters from the Voua river. Meadows are dominated by *Halophila ovalis* ssp. *bullosa*, *Halodule uninervis* with some *Halodule pinifolia*. *H. uninervis* is much denser in the channels or intertidal pools. Two Seagrass-Watch sites (NN1, NN2) are located on intertidal banks separating Cuvu village and Shangri-la Fijian Resort. The sites are monitored by Alfred Ralifo, Nadroga Navosa Provincial High School students and Seagrass-Watch HQ. Mounded topography formed by callianassid shrimp is ubiquitous on the sandy carbonate intertidal banks. At low tide, villagers fish and glean the intertidal flats.

Seagrasses are also found on the inshore area from the Naidiri River to Nakorola Point at **Malomalo**. The Malomalo seagrass meadows are dominated by *Halodule uninervis* (Solomona *et al.* 2002). Their interlacing rhizome / root mat providing stability to the sand and rubble zone prevalent in this area. They also function as a nursery for organisms, although crab holes and the synaptid *S. maculata*, were the only form of species presence recorded in these meadows.

In the heart of the Coral Coast is **Tagaqe** where a Seagrass-Watch site (TQ1) is established in the 1.6 hectare *Halodule pinifolia* dominate meadow. The site is on the intertidal reef-flat in front of Hideaway Resort, and is monitored by Seagrass-Watch HQ. The site is also immediately adjacent to a “*tabu*” area declared by Tagaqe village. Just over 20% of the seagrass meadow is within the designated *tabu* area. Four seagrass species are found in the meadow: *Halodule uninervis*, *Halodule pinifolia*, *Halophila ovalis* ssp. *bullosa* and *Syringodium isoetifolium*. The most noticeable feature of the meadow condition is the high amount of epiphyte cover on the leaves, possibly a consequence of elevated water column nutrients.



Suva

Nukubocu is part of a barrier reef system which encloses Laucala Bay and Suva harbour, in the Suva Peninsula. Seagrasses are widespread in the back reef regions, and dense *Syringodium isoetifolium* meadows fringe the channels between the reefs. Until recently, coral sand was also dredged from the back reef region of Nukubocu reef by a local industry as raw material in the cement making process. *Halodule uninervis*, *Halodule pinifolia* and *Halophila ovalis* ssp. *bullosa* are also found on the reef flat and back reef, often in a mosaic of patchy meadows

Seagrass meadows also cover much on the intertidal reef platform surrounding Suva Point and surrounding the shores of **Laucala Bay**. Seagrass-Watch has one site (SV1) established on the intertidal banks at **Nasese**. The meadow is comprised of *Halodule uninervis*, *Halodule pinifolia* and *Halophila ovalis* ssp. *bullosa*. Of concern, are the high amounts of epiphytic algae covering the leaves and macroalgae, which formed a thick mat over the grass.

Tailevu (Bau landing to Natovi)

The area north east of Suva has patch reef and fringing reef complexes, quite unique in the south Pacific. Shallow subtidal areas of *Halodule uninervis*, *Halophila ovalis*, and *Syringpdium isoetifolium* are reported to be found in the area, although no detailed survey has been conducted (Nair et al. 2006).

LOMAIVITI GROUP

Seagrass meadows are scattered throughout the Lomaiviti Group, where they are recognised as significant areas for green turtle foraging.

Ovalau

Significant seagrass meadows are present on the intertidal fringing reefs and within the lagoons surrounding Ovalau. Dense meadows of *Syringpdium isoetifolium* are located inside the barrier reef on either side of the entrance to **Levuka Harbour**. A narrow band of *Halophila ovalis* separates the dense *Syringpdium isoetifolium* meadows from the back reef, and isolated patches of *Halodule uninervis* are scattered throughout.

Sparse *Halodule pinifolia* and *Halophila ovalis* ssp. *bullosa* meadows are scattered across the intertidal fringing reefs between **Nacobo** and Cawaci (incl. Levuka seafront and **Vagadaci** Bay). Meadows are denser and more diverse adjacent to mangrove areas. Large intertidal meadows of *Halodule uninervis*, *Halodule pinifolia*, *Syringodium isoetifolium* and *Halophila ovalis* ssp. *bullosa* are found on the fringing reef opposite St John's College at **Cawaci**. Two Seagrass-Watch sites (CW1, CW2) are located in front of St John's College, monitored by Masao Yoshida, Shaun Ashley, Charlene Ashley and Seagrass-Watch HQ. In recent years the reef has experienced blooms of green algae and physical disturbance from PWD extraction activities. The fringing reef is popular at low tide with villagers fishing and gleaning.

Gau Island

Seagrass meadows have been reported on the fringing reef platforms of Gau, adjacent to Lovu and Vadravadr villages. The shoreline of Tikina Sawaieke presents a series of extensive seagrass meadows located near mangroves and mudflats. Two seagrass species (*Halodule uninervis* and *Halodule ovalis*) are found in meadows where *Halodule uninervis* is the dominate species closer to shore, with higher cover, decreasing seaward where *Halodule ovalis* become more dominant (Fiu 2005). Overall, seagrass meadows in Tikina Sawaieke are relatively healthy in terms of the



extensive growth to at least 0.5 km from shore. Villagers have noticed the regrowth of the seagrass in their local shore area as a healthy indication of the marine environment; however these areas are being impacted from boating activity.

Kadavu

Significant *Syringodium isoetifolium* meadows have been reported and studied in the lagoon at **Dravuni**. *Halodule uninervis*, *Halodule pinifolia* and *Halophila ovalis* ssp *bullosa* are also present in varying amounts. Seagrass meadows are also reported from the south eastern coasts of **Kadavu** Island (e.g., adjacent to Matava Resort), although no species are described. As the islands are protected by the Great Astrolabe Reef, the longest barrier reef in Fiji, it is possible that extensive subtidal meadows may exist.

VANUA LEVU

Until recently little was known about the northern shores of the second largest island in Fiji, Vanua Levu, in particular Cakaulevu (literally the 'Big Reef') or the Great Sea Reef. Running parallel to the coastline of the provinces of Macuata and Bua, the **Great Sea Reef** area (including barrier reef with inshore mangrove islands and fringing reefs) was identified as a globally significant area by a variety of stakeholders at the FIME vision workshop in December 2003 (Heaps 2005; Nair *et al.* 2006). In December 2004, a biological survey expedition of the area further identified significant seagrass meadows (*Halodule uninervis*, *Halodule pinifolia*, *Halophila ovalis*, *Halophila ovalis* ssp *bullosa* and *Syringodium isoetifolium*) surrounding the coral/mangrove islets of Vatuka. During this expedition, the first collection of *Halophila decipiens* was recorded for Fiji (Skelton and South 2006).

To the north east, there have been anecdotal reports of *Halodule uninervis* and *Halophila ovalis* on fringing reefs in the **Nabuna** area and on the mudflats and slopes within **Natewa Bay** (Nair *et al.* 2006).

On the southern shores of Vanua Levu, seagrasses appear to be restricted to bays which provide some protection from the prevailing weather. Significant areas of seagrass (*Halodule uninervis*, *Halodule pinifolia*, *Halophila ovalis* ssp *bullosa* and *Syringodium isoetifolium*) have been reported in the past in **Wainsunu** Bay and on the scattered reefs of **Savusavu** Bay, although these appear to have declined in recent years (Nair *et al.* 2006).

LAU GROUP

Sparse to moderately abundant *Halodule uninervis* and *Halophila ovalis* meadows are common on reef platforms within the group. On **Kabara**, seagrass meadows are located on both leeward and windward sides (Fiu 2004). The meadows are composed of *Halophila ovalis* and *Halodule uninervis* and abundance is high. Seagrass meadows on the leeward (protected) side of the island (e.g., adjacent to Naikeleaga) are higher in abundance with more macroalgae, than compared to the windward meadows (e.g., adjacent to Udu). Invertebrates (including urchins, sea hares and clams) and turtles however, were higher in windward meadows than leeward.

Fulaga in southern Lau has an interesting outlay of limestone islets which creates marine caves, fringing and atoll reefs and lagoons which host seagrass meadows with *Halodule* species dominant (Nair *et al.* 2006).

YASAWA GROUP

In the Yasawa Group, seagrass has been reported from the fringing reefs and the lee slopes of islands (Nair *et al.* 2006). Unidentified seagrass has been described in patches on white, coarse, coronous sand adjacent to Malakati Village, **Nacula** Island (Parkinson 1982). *Halophila* spp. have also been reported from the subtidal slopes in protected bays of the island group (V. Vuki Pers. Comm.). Unfortunately detailed information is not readily available, although it can possibly be gleaned from consultancy reports and environmental impacts assessments associated with proposed developments in the region.

MAMANUCA GROUP

The Mamanuca Islands in western Fiji have been the focus of tourism development in Fiji for many years and the industry is very much aware of the value of conserving the reef habitats and fostering sustainable development.

During a survey to characterise the major benthic marine habitats of five reef complexes in the Mamanuca Islands, baseline transects were examined at **Mana** Island, **Navini** Island, the **Namotu** and **Malolo** Island groups (Harborne *et al.* 2001). From a preliminary habitat map produced using 'Landsat 7' satellite imagery, coupled with ground truthing characterisation, the area occupied by each of the habitats within the project area (1826 km²) was calculated. Only one of the seven major benthic classes included seagrass and it was estimated to cover 6.46 km². As the habitat discrimination was no more detailed than "sand with sparse algae and seagrass", the species of seagrass present were not identified, but possibly include *H. pinifolia* and *H. ovalis*, similar to nearby Castaway Island

During baseline surveys to characterise the marine habitats adjacent to **Castaway** Island, Solandt *et al.* (2002) identified sparse *Halophila ovalis* and *Halodule pinifolia* meadows from 4 transects on the fringing reefs and in the shallow subtidal waters.

No other information is available on seagrasses of the Mamanuca Islands, however large meadows of *Syringodium isoetifolium*, *Halophila ovalis* and *Halodule pinifolia* are known to fringe the adjacent Viti Levu coast surrounding **Denarau** Island (e.g., Sofitel and Sheraton Resorts) (Pers. Obs.).



L: *Syringodium isoetifolium* meadow, C: *Halodule pinifolia* meadow, R: *Halophila ovalis* ssp. *bulbosa* meadow
Photos: Denarau, Viti Levu, Jan 2007, by Len McKenzie

ROTUMA

Only one seagrass species has been reported from Rotuma: *Syringodium isoetifolium*. In late 2004 and 2005, Laje Rotuma conducted seagrass monitoring in the *Syringodium isoetifolium* in **Motusa** Bay (Alfred Ralifo, Pers. Comm.). The results showed a percentage seagrass cover increased by 6% from 2004 to 2005, indicating that the seagrass was relatively healthy. There was also a reduction in the percentage epiphytic and macro-algae. The mean number of animals found on the seagrass meadow also increased slightly from 2004 to 2005. Seagrass has also been reported in **Maka** Bay (Nair *et al.* 2006).

Studies on seagrasses in Fiji are limited. The only detailed studies of biological processes of seagrass meadows have been conducted at Dravuni Island: nutrient dynamics and carbon/nitrogen (Yamamuro *et al.* 1993); growth and production (Aioi and Pollard 1991); irradiance/productivity (Pollard and Aioi 1991); litter production and decomposition (Pollard and Kogure 1993).

The total area of seagrasses world-wide is estimate to be at least 177,000 sq km (Spalding *et al.* 2003). The total area of seagrass meadows in the Fiji Islands however is unknown, as no broad scale mapping exercise has been conducted (Coles *et al.* 2003). This is because mapping in tropical systems is generally from field observations as remotely sensed data (satellite and aerial imagery) is often ineffective for detecting tropical seagrasses of low biomass and/or in turbid water (McKenzie *et al.* 2001). New technologies may assist and there are several projects currently underway attempting to at map Fiji's seagrass resources, mainly at the local level.

IMPACTS

Seagrasses in Fiji are threatened by a number of impacts, including tourism, improper methods of disposal of solid waste, sewage pollution, depletion of fisheries, coral harvesting, coastal erosion, storm surge and flooding, siltation of rivers and coastal areas as a result of soil erosion inland agriculture and forestry and sand mining. Long-term ecological studies of seagrass meadows on Suva Reef revealed that losses occurred in some years because of major disturbances such as tsunamis, cyclones and flood (Vuki 1994). Analysis of spatial pattern of seagrass meadows from airborne images showed clearly that there were oscillations in abundance on Suva Reef; seagrass meadows extended towards the lagoon in some years and regressed in others. Regressions in seagrass meadows on Suva back reef areas were attributed to high turbidity and siltation cause by foreshore reclamations (Vuki 1994). Coastal modifications mostly occur near major urban areas and coastal towns. At least 110 hectares have been reclaimed in Suva since 1881 (Vuki et al 2000)

All of these issues and associated threats pose a challenge for conserving healthy seagrass meadows in the Fiji Islands.



Above left: *Halodule uninervis* & *Halophila ovalis* ssp. *bullosa*, Nadroga Navosa, Viti Levu, Jan 2007, photo by Len McKenzie
 Above right: *Halodule pinnifolia* meadow, Natadola, Viti Levu, April 2006, photo by Len McKenzie
 Below: *Halophila ovalis* ssp. *bullosa*, Korotogo, Viti Levu, April 2006, photo by Len McKenzie



A guide to the identification of seagrasses in the Fiji Islands

Adapted from Waycott, M, McMahon, K, Mellors, J., Calladine, A., and Kleine, D (2004) A guide to tropical seagrasses in the Indo-West Pacific. (James Cook University Townsville) 72pp.

Leaves cylindrical



cylindrical

Syringodium isoetifolium

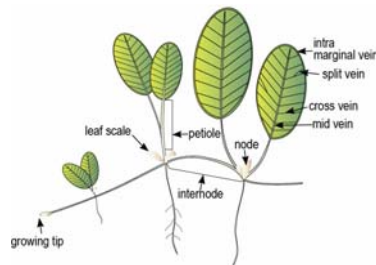
- Leaf tip pointed
- Leaves contain air cavities
- Inflorescence a “cyme”

Leaves oval to oblong



oval to oblong

leaves with petioles, in pairs



Halophila ovalis

- cross veins more than 10 pairs
- leaf margins smooth
- leaf surface smooth
- no leaf hairs
- separate male & female plants

Halophila ovalis ssp *bullosa*

- leaf surface bullate (blister or pucker-like)

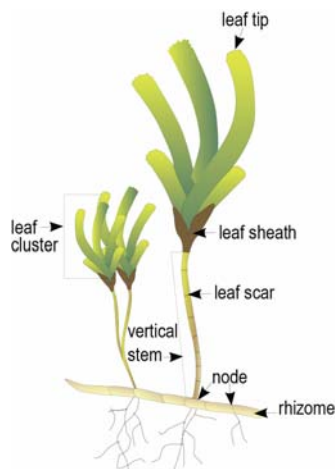
Halophila decipiens

- leaf margins serrated
- fine hairs on both sides of leaf blade
- male & female flowers on same plant

Leaves strap-like



strap-like



Halodule uninervis

- Leaf tip tri-dentate or pointed, not rounded
- Leaf with 3 distinct parallel- veins, sheaths fibrous
- Rhizome usually white with small black fibres at the nodes

Halodule pinifolia

- Leaf tip rounded
- Leaf with 3 distinct parallel- veins, sheaths fibrous
- Rhizome usually white with small black fibres at the nodes

Monitoring a seagrass meadow

Environment monitoring programs provide coastal managers with information and assist them to make decisions with greater confidence. Seagrasses are often at the downstream end of catchments, receiving runoff from a range of agricultural, urban and industrial land-uses.

Seagrass communities are generally susceptible to changes in water quality and environmental quality that make them a useful indicator of environmental health. Several factors are important for the persistence of healthy seagrass meadows, these include: sediment quality and depth; water quality (temperature, salinity, clarity); current and hydrodynamic processes; and species interactions (e.g., epiphytes and grazers). Seagrass generally respond in a typical manner that allows them to be measured and monitored. In reporting on the health of seagrasses it is important to consider the type of factors that can effect growth and survival. Factors include:

- increased turbidity reduces light penetration through the water, interfering with photosynthesis and limiting the depth range of seagrass;
- increased nutrient loads encourages algal blooms and epiphytic algae to grow to a point where it smothers or shades seagrasses, also reducing photosynthetic capacity;
- increased sedimentation can smother seagrass or interfere with photosynthesis;
- herbicides can kill seagrass and some chemicals (e.g., pesticides) can kill associated macrofauna;
- boating activity (propellers, mooring, anchors) can physically damage seagrass meadows, from shredding leaves to complete removal;
- storms, floods and wave action can rip out patches of seagrasses.

Seagrass-Watch

A simple method for monitoring seagrass resources is used in the Seagrass-Watch program. This method uses 50m by 50m sites established within representative intertidal meadows to monitor seagrass condition. The number and position of sites can be used to investigate natural and anthropogenic impacts.

Seagrass-Watch is one of the largest seagrass monitoring programs in the world. Since it's genesis in 1998 in Australia, Seagrass-Watch has now expanded internationally to 18 countries. Monitoring is currently occurring at over 165 sites. To learn more about the program, visit www.seagrasswatch.org.

Seagrass-Watch aims to raise awareness on the condition and trend of nearshore seagrass ecosystems and provide an early warning of major coastal environment changes. Participants of Seagrass-Watch are generally volunteers from a wide variety of backgrounds who all share the common interest in marine conservation and sustainability. Most participants are associated with established local community groups, schools, universities & research institutions, government (local & state) or non-government organisations.

Seagrass-Watch integrates with existing education, government, non-government and scientific programs to raise public awareness of these important marine ecosystems for the benefit of the community. The program has a strong scientific underpinning with an emphasis on consistent data collection, recording and reporting. Seagrass-Watch identifies areas important for seagrass species diversity and sustainability and the information collected is used to assist the management of coastal environments and to

prevent significant areas and species being lost. Seagrass-Watch is also a component of the Global Seagrass Monitoring Network.

Seagrass-Watch monitoring efforts are vital to assist with tracking global patterns in seagrass health, and assess the human impacts on seagrass meadows, which have the potential to destroy or degrade these coastal ecosystems and decrease their yield of natural resources. Responsive management based on adequate information will help to prevent any further significant areas and species being lost. To protect the valuable seagrass meadows along our coasts, the community, government and researchers have to work together.

THE GOALS OF THE PROGRAM ARE:

- To educate the wider community on the importance of seagrass resources
- To raise community awareness of coastal management issues
- To develop community participation and ownership
- To build the capacity of local communities in the use of standardised scientific methodologies
- To conduct long-term & broad-scale monitoring of seagrass & coastal habitats, and
- To provide an early warning system of coastal environment changes for management.

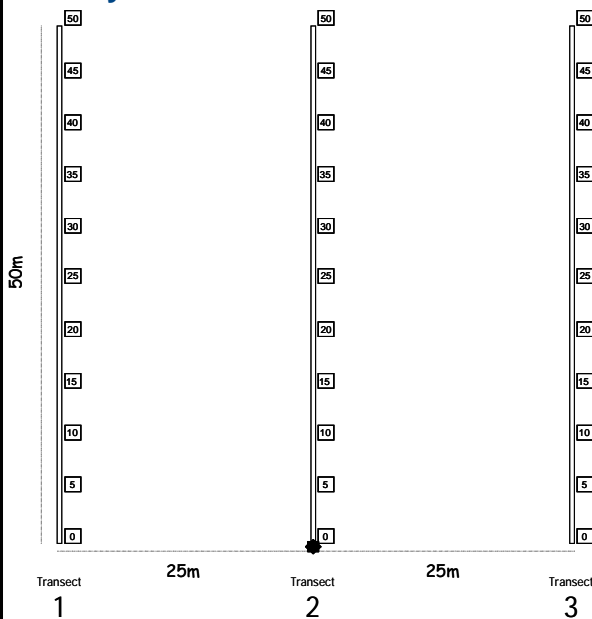


Levuka reef, Ovalau, Jan 2007, photo by Len McKenzie

Seagrass-Watch Monitoring Summary

Source: McKenzie, L.J., Campbell, S.J. & Roder, C.A. (2003) *Seagrass-Watch: Manual for Mapping & Monitoring Seagrass Resources by Community (citizen) volunteers.* (DPI&F, NFC, Cairns) 100pp (www.seagrasswatch.org/manuals.html)

Site layout



Quadrat code = site + transect+quadrat

e.g., CW1225 = Cawaci site 1, transect 2, 25m quadrat

Pre-monitoring preparation

Make a Timetable

Create a timetable of times of departure and arrival back, and what the objective of the day is and what is to be achieved on the day. Give a copy of this to all volunteers involved in advance so they can make their arrangements to get to the site on time. List on this timetable what the volunteers need to bring.

Have a Contact Person

Arrange to have a reliable contact person to raise the alert if you and the team are not back at a specified or reasonable time.

Safety

- Assess the risks before monitoring - check weather, tides, time of day, etc.
- Use your instincts - if you do not feel safe then abandon sampling.
- Do not put yourself or others at risk.
- Wear appropriate clothing and footwear.
- Be sun-smart.
- Adult supervision is required if children are involved
- Be aware of dangerous marine animals.
- Have a first aid kit on site or nearby
- Take a mobile phone or marine radio

Necessary equipment and materials

- | | |
|--|---|
| <input type="checkbox"/> 3x 50metre fibreglass measuring tapes | <input type="checkbox"/> Clipboard, pencils & 30 cm ruler |
| <input type="checkbox"/> 6x 50cm plastic tent pegs | <input type="checkbox"/> Camera & film |
| <input type="checkbox"/> Compass | <input type="checkbox"/> Quadrat photo labeller |
| <input type="checkbox"/> 1x standard (50cm x 50cm) quadrat | <input type="checkbox"/> Percent cover standard sheet |
| <input type="checkbox"/> Magnifying glass | <input type="checkbox"/> Seagrass identification sheets |
| <input type="checkbox"/> 3x Monitoring datasheets | |

Quarterly sampling

Within the 50m by 50m site, lay out the three 50 transects parallel to each other, 25m apart and perpendicular to shore (see site layout). Within each of the quadrats placed for sampling, complete the following steps:

Step 1. Take a Photograph of the quadrat

- Photographs are usually taken at the 5m, 25m and 45m quadrats along each transect, or of quadrats of particular interest. First place the photo quadrat labeller beside the quadrat and tape measure with the correct code on it.
- Take the photograph from an angle as **vertical** as possible, which includes the entire quadrat frame, quadrat label and tape measure. Avoid having any shadows or patches of reflection off any water in the field of view. Check the photo taken box on datasheet for quadrat.

Step 2. Describe sediment composition

- Dig your fingers into the top centimetre of the substrate and feel the texture. Describe the sediment by noting the grain size in order of dominance (e.g., Sand, Fine sand, Fine sand/Mud).

Step 3. Estimate seagrass percent cover

- Estimate the total % cover of seagrass within the quadrat – use the percent cover photo standards as a guide.

Step 4. Estimate seagrass species composition

- Identify the species of seagrass within the quadrat and determine the percent contribution of each species to the cover. Use seagrass species identification keys provided.

Step 5. Measure canopy height

- Measure canopy height of the seagrass ignoring the tallest 20% of leaves. Measure from the sediment to the leaf tip of at least 5 shoots.

Step 7. Estimate algae percent cover

- Estimate % cover of algae in the quadrat. Algae are seaweeds that may cover or overlie the seagrass blades. Use "Algal percentage cover photo guide".

Step 8. Estimate epiphyte percent cover

- Epiphytes are algae attached to seagrass blades and often give the blade a furry appearance. First estimate how much of the blade surface is covered, and then how many of the blades in the quadrat are covered (e.g., if 20% of the blades are each 50% covered by epiphytes, then quadrat epiphyte cover is 10%).

Step 9. Describe other features and ID/count of macrofauna

- Note and count any other features which may be of interest (eg. number of shellfish, sea cucumbers, sea urchins, evidence of turtle feeding).

Step 10. Take a voucher seagrass specimen if required

- Seagrass samples should be placed inside a labelled plastic bag with seawater and a waterproof label. Select a representative specimen of the species and ensure that you have all the plant part including the rhizomes and roots. Collect plants with fruits and flowers structures if possible.

At completion of monitoring

Step 1. Check data sheets are filled in fully.

- Ensure that your name, the date and site/quadrat details are clearly recorded on the datasheet. Also record the number of other observers assisting.

Step 2. Remove equipment from site

- Remove all tent pegs and roll up the tape measures. If the tape measures are covered in sand or mud, roll them back up in water.

Step 3. Wash & pack gear

- Rinse all tapes, pegs and quadrats with freshwater and let them dry.
- Review supplies for next quarterly sampling and request new materials
- Store gear for next quarterly sampling

Step 4. Press any voucher seagrass specimens if collected

- The voucher specimen should be pressed as soon as possible after collection. Do not refrigerate longer than 2 days, press the sample as soon as possible.
- Allow to dry in a dry/warm/dark place for a minimum of two weeks. For best results, replace the newspaper after 2-3 days.

Step 5. Submit all data

- Data can be entered into the MS-Excel file downloadable from www.seagrasswatch.org. Email completed files to hq@seagrasswatch.org
- Mail original datasheets, photos and herbarium sheets

Seagrass-Watch HQ
Northern Fisheries Centre
PO Box 5396
Cairns QLD 4870 AUSTRALIA



Seagrass mapping

Seagrass-Watch activities initially map the distribution of seagrass meadows at a locality or in a region to better understand the seagrass resources of an area. Mapping is often limited to the accessible intertidal seagrasses, although in some cases subtidal seagrass meadows can be included.

The most important information that is required for management of seagrass resources is their distribution, ie. a map. It would be inappropriate to set up a monitoring program if the most basic information is unavailable - that is, whether seagrass is present or absent.

When planning a mapping task, there are several issues that need to be considered, including:

Scale

Mapping requires different approaches depending on whether survey area is relative to a region (tens of kilometres), locality (tens of metres to kilometres) or to a specific site (metres to tens of metres). Scale includes aspects both of extent and resolution. In both broad and large scale approaches, the intensity of sampling will be low (low resolution), with a statistical sampling design that allows the results to be extrapolated from a few observations to the extent of the study area. For finer scale examinations of seagrass meadows, the sampling intensity required can be high with greater precision (high resolution). Scale also influences what is possible with a limited set of financial and human resources. The financial, technical, and human resources available to conduct the study is also a consideration.

Accuracy

Determining the level of detail required when mapping an area also depends on the level of accuracy required for the final map product. Errors that can occur in the field directly influence the quality of the data. It is important to document these. GPS is a quick method for position fixing during mapping and reduces point errors to <3m in most cases. It is important for the observer to be as close as possible to the GPS aerial receiver to minimise position fix error.

Choosing a Survey/Mapping strategy

The selection of a mapping scale represents a compromise between two components. One is the maximum amount of detail required to capture the necessary information about a resource. The other is the logistical resource available to capture that level of detail over a given area. Generally, an area can be mapped using a grid pattern or a combination of transects and spots. When mapping a region of relatively homogenous coastline between 10 and 100 km long, we recommend that transects should be no further than 500-1000 m apart. For regions between 1 and 10 km, we recommend transects 100-500 m apart and for localities less than 1 km, we recommend 50-100 m apart. This however may change depending on the complexity of the regional coastline, i.e., more complex, then more transects required.

To assist with choosing a mapping strategy, it is a good idea to conduct a reconnaissance survey. An initial visual (reconnaissance) survey of the region/area will give you an idea as to the amount of variation or patchiness there is within the seagrass meadow. This will influence how to space your ground truthing sites.

When mapping, ground truthing observations need to be taken at regular intervals (usually 50 to 100m apart). The location of each observation is referred to a point, and the intervals they are taken at may vary depending on the topography.

When ground truthing a point, there are a variety of techniques that can be used depending on resources available and water depth (free dives, grabs, remote video, etc). First the position of a point must be recorded, preferably using a GPS. Otherwise use a handheld compass to determine the bearing, triangulating to at least 3 permanent landmarks or marker established as reference points. A point can vary in size depending on the extent of the region being mapped. In most cases a point can be defined as an area encompassing a 5m radius. Although only one observation (sample) is necessary at a ground truth point, we recommend replicate samples spread within the point (possible 3 observations) to ensure the point is well represented.

Observations recorded at a point should ideally include some measure of abundance (at least a visual estimate of biomass or % cover) and species composition. Also record the depth of each point (this can be later converted to depth below mean sea level) and other characteristics such as a description of the sediment type (eg. shell grit, rock, gravel, coarse sand, sand, fine sand or mud), or distance from other habitats (reefs or mangroves).

For details on the necessary materials & equipment and the general field procedure for mapping a seagrass meadow, please see Chapter 4 of the Seagrass-Watch manual (www.seagrasswatch.org/manuals.html).

Creating the map

The simplest way to map the distribution of seagrasses is to draw the meadows on a paper marine chart from the GPS positions of the ground truth sites. The problem with this type of mapping is that the final map is in a format that does not allow manipulation and transformation. The layout of a paper map is permanent, which makes it difficult for future seagrass mapping studies to be compared, queried and analysed. If resources are available, we recommend that the data be transferred to a digital format and a Geographic Information System (GIS) be used. A detailed description of using and mapping with GIS is beyond the scope of this manual, and we recommend consulting with a scientist experienced in mapping and reading McKenzie *et al.* (2001b).



Using Global Positioning System (GPS) to map the meadow edge/boundary

The meadow boundary is mapped by recording (or marking) a series of waypoints along the edge of the meadow from which a line representing the meadow boundary can then be drawn.

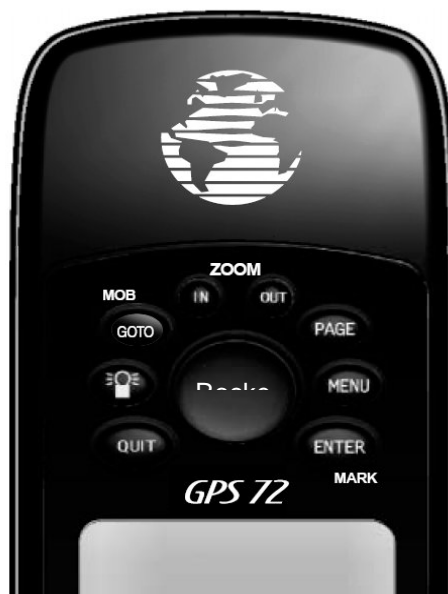
To map the boundary you should:

- Walk along the meadow edge, stopping to record (mark) waypoints on your GPS approximately every 10 steps or where there is a change in direction of the meadow boundary.
- Alternatively, you can use the “track” feature if the meadow boundary is clear.
- Only record a waypoint when you are sure you are on the edge of the meadow. For meadows where the edge is patchy you may have to do some reconnaissance to make sure you are at the edge of the seagrass meadow before recording a waypoint.
- Keep in mind that others may be using the points that you record on the GPS to draw a line (*“join the dots”*) that represents the meadow boundary, so it is better to have too many points than too few.
- Stand still for approximately 10 seconds on a point before recording (marking) a waypoint. It is good to keep checking the accuracy (in the top right of the screen) to ensure the point is as accurate as possible (generally less than 5m).
- As waypoints are labelled by sequential numbers, ensure you record the label of the first and last waypoint. If you make an error, either delete the waypoint or note the erroneous waypoint.
- At the conclusion of the sampling trip the waypoints you record should be downloaded onto your computer (see later section)

Using the GARMIN GPS 72

Turn GPS Receiver On.

Hold down the Red button for 2 seconds. Wait a few seconds and the GPS unit will begin to track available satellites. Make sure to hold the GPS in a vertical position (top to the GPS pointing to the sky) for best reception of satellite signals.

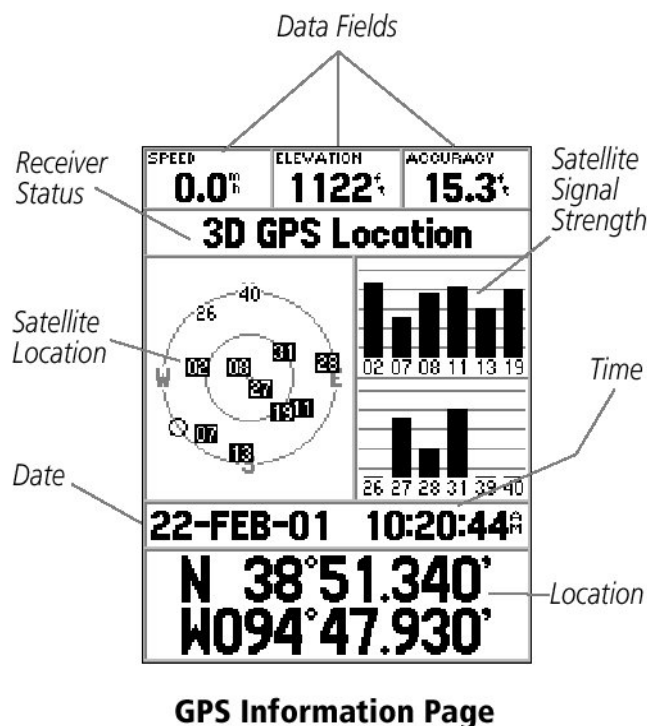


Using the Interface Keys on the GPS

- **IN & OUT** keys – used on the Map Page to zoom in and out.
- **GOTO/MOB** key – used to begin or stop navigation to a waypoint.
- **PAGE** key – used to cycle through the five main display pages in sequence.
- **POWER** key – used to turn the GPS on or off by holding the button down for 2 seconds.
- **MENU** key – used to display a variety of GPS options which can be altered to suit usage. Press the Menu key twice to access the Main Menu.
- **QUIT** key – used to cycle in reverse to the Page key. Also used to cancel operation in progress.
- **ENTER/MARK** key – used to activate or confirm a selection. To record a Waypoint, press and hold down the ENTER/MARK key.
- **ROCKER** key – used to move up/down and left/right to access and change menu options, and for data entry. The ROCKER key is always used to scroll through the different menu options and the ENTER key is used to activate or confirm the selection.

Tracking satellites.

Continue to press the PAGE key until the GPS Information Page appear (the Information Page looks like the diagram just below). When the GPS receiver is tracking enough satellites (minimum four), the GPS Information appears.



Receiver is now ready to record Waypoints. It is recommended that there are at least four satellites are present and have a strong signal when marking the waypoints. The more satellites available the better the accuracy.

Before any waypoints are to be collected, you must check that the GPS settings are correct. The settings are in the Main Menu.



Setting up the correct properties in the Main Menu

- To get to the Main Menu you must press the **Menu** key twice.
- Scroll down to highlight “Setup” and press **Enter**.
 1. You must use the **ROCKER** key and move to the appropriate field and use the **ENTER** key to activate the field.
 2. Then use the **ROCKER** key (to move up/down and left/right) to access the appropriate setting and use the **ENTER** key to confirm selection.
 3. Once changes are made, use the **QUIT** to exit each of the option or menu.
- Scroll across to the “Time” tab and change to the Time settings.

Time Setup

Time Format : 24 Hour

Time Zone : Other

UTC Offset: +10:00

- Scroll across to the “Location” tab and change to the Time settings.

Units Setup

Elevation : Meters

Distance and Speed: Metric

Temperature: Celsius

- Scroll across to the “Location” tab and change to the Time settings.

Location Setup

Location Format : hddd.ddddd

MAP DATUM : WGS84

- Scroll across to the “Interface” tab and change to the Time settings.

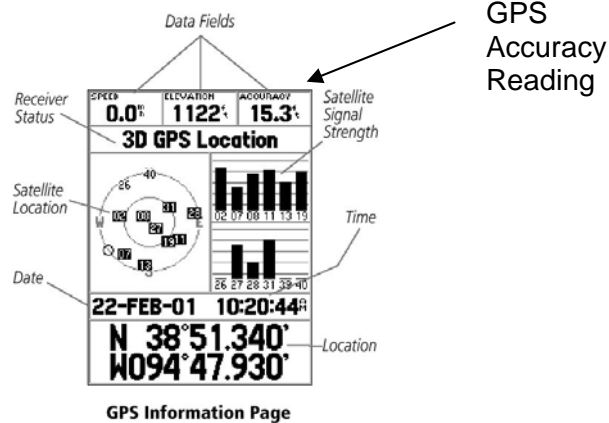
Interface Setup

Serial Data Format: Garmin

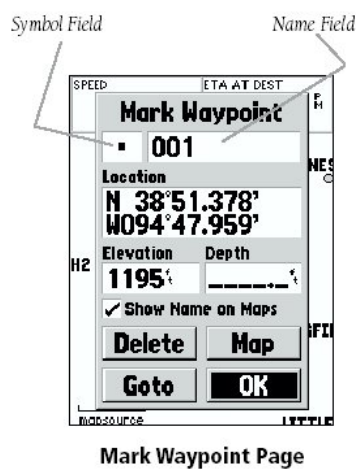
Mark Location as Waypoints

Mark Waypoints are stored as point locations.

To record Waypoints, it is best to be in the Information Page (continue to press the PAGE button until the Information page appears). Before you record a Waypoint, you must look at the accuracy reading on the GPS Information Page. An acceptable accuracy should be less than 15 metres (<15 m)



Step 1. Press and hold the **ENTER** key. The following screen appears:



To save the waypoint without any changes, press **ENTER** on the OK button to save the waypoint.

- Step 2.** You can choose to rename the waypoint number to another name by using the **ROCKER** key to highlight the Name field, then press **ENTER**.
- Step 3.** Use the **ROCKER** key as an Alpha-numeric key selection to type in the new waypoint name. Press the **ENTER** key to accept the name.
- Step 4.** You can choose to delete the waypoint by highlighting 'DELETE?', press **ENTER** and then highlight the 'Yes' prompt and press **ENTER** again to confirm.

Use the Goto option to get to a location.

- Press the **GOTO** key.
- Highlight the "Waypoint" then press the **ENTER** key.
- Use the **ROCKER** key to highlight the waypoint you want to go to, press the **ENTER** key.
- Press the **PAGE** key until you get to the Map Page to see where you are (use the **IN** or **OUT** key to zoom in or out). Press the **PAGE** key until you get to the Compass Page to see where you need to be heading to get to the waypoint.



Installing the program MAPSOURCE to download information from your GPS to your computer

Step 1: Installing MapSource

Before installing MapSource, all programs should be closed.

1. Put CD into the CD Drive (Place the CD in with CD labels facing up)
2. If the MapSource setup menu does not appear automatically, Go to **Windows Explorer** (Select Start, Programs, Windows Explorer)
 - Double Click on CD Drive (This could either be D: or E: drive on your computer)
 - Double click on "**Setup.exe**" (To startup the setup program [about 70kb])
 - i. Click **Next** (To say OK to the Welcome Page)
 - ii. Click **Yes** (To accept License Agreement)
 - iii. Click **No** (When ask to check GPS)
 - iv. Click **No** (To confirm not checking the GPS)
 - v. Click **Next** (To select typical install)
 - vi. Click **Finish** (To complete installation)

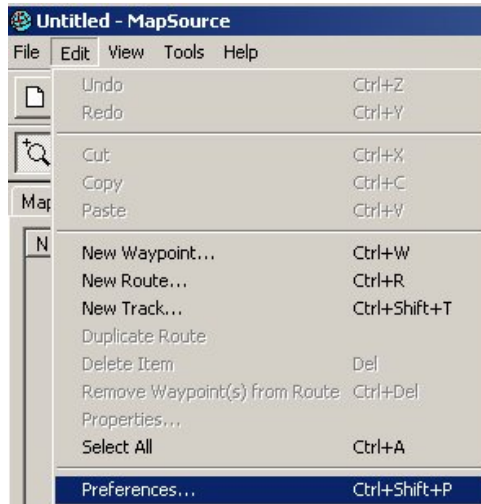
Step 2: Installing Updated files for Mapsource

1. While the **CD** is still in the CD drive,
3. Go to **Windows Explorer** (Select Start, Programs, Windows Explorer)
2. Double Click on CD Drive (This could either be D: or E: drive on your computer)
3. Double Click the **Mapsource Upgrade** folder (You will see a MapSource_680.exe file)
4. Double Click on the file **mapsource_680.exe** (This will startup the setup program to MapSource)
5. Click **Next** (To choose English language)
6. Click **Next** (To accept MapSource Version 6.8)
7. Click **Yes** (To accept License Agreement)
8. Click **Finish** (To complete installation)
9. Eject CD and take disc out

Step 3: Setting the correct Properties for MapSource

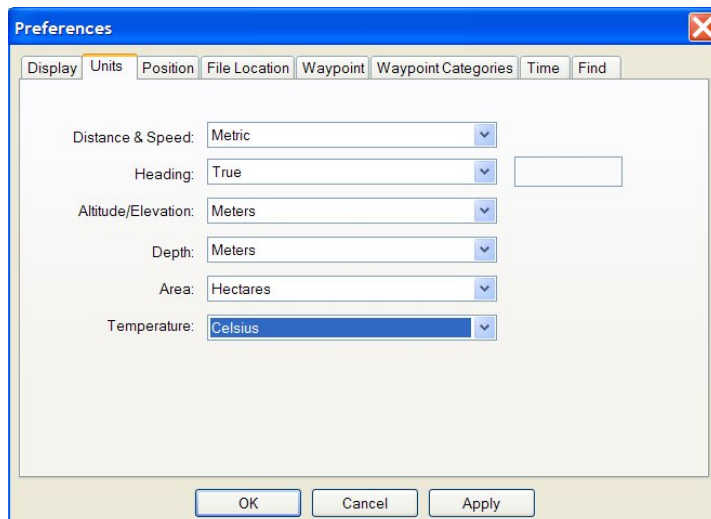
Before using MapSource you must set up some map properties to view the data.

1. Start MapSource (Click the MapSource Icon on the desktop)
2. Go to the **Edit** menu and select **Preferences**



3. The Preferences window will appear.

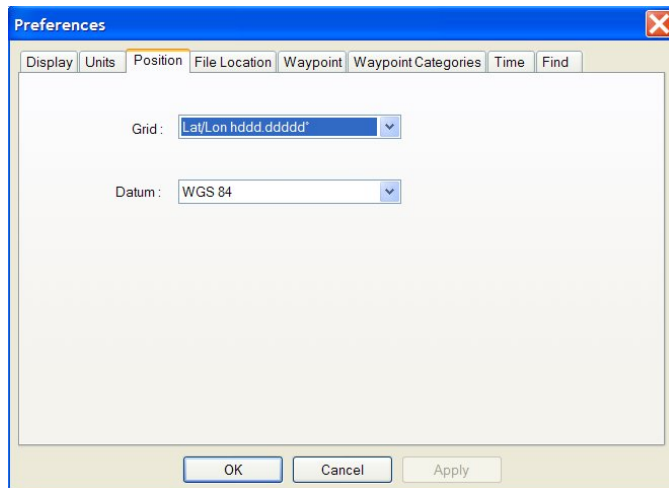
Go to the **Units** tab



You must set the following properties:

- Distance & Speed Metric
- Heading True
- Altitude/Elevation Meters
- Depth Meters
- Area Hectares
- Temperature Celsius

4. Go to the **Position** tab



You must set the following properties:

- Grid Lat/Lon hddd.ddddd
- Datum WGS 84

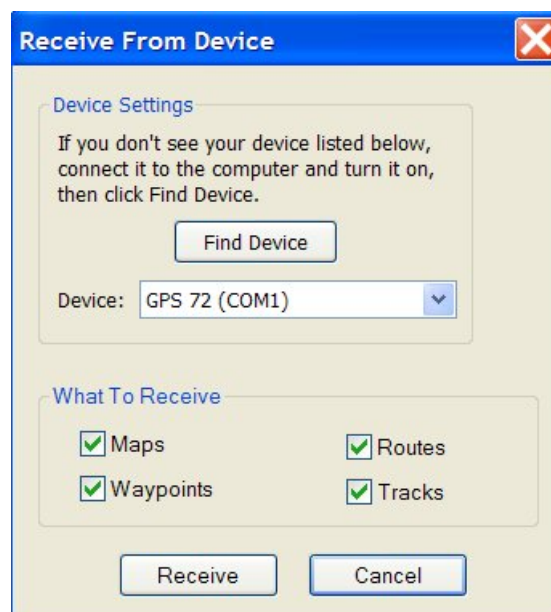
5. Click OK to finish.

6. MapSource is now correctly set up for use.

Downloading information from your GPS using the MapSource program

Step 1: Downloading GPS data using MapSource (download software)

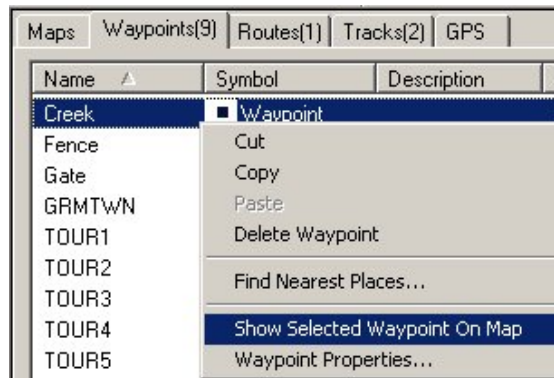
1. Start MapSource (Double click on the MapSource Icon) or (start, programs, MapSource, MapSource)
2. Plug the Garmin serial cable from your GPS unit to the communications port on your computer.
3. On the GPS unit, make sure that the Garmin interface is set to “**GARMIN**” on the GPS 72. To Check this, go to the “Setup menu”, select “Interface”, select “GARMIN”.
4. In the MapSource program, go to the **Transfer** menu and select “**Receive from Device..**”. The following menu will appear.



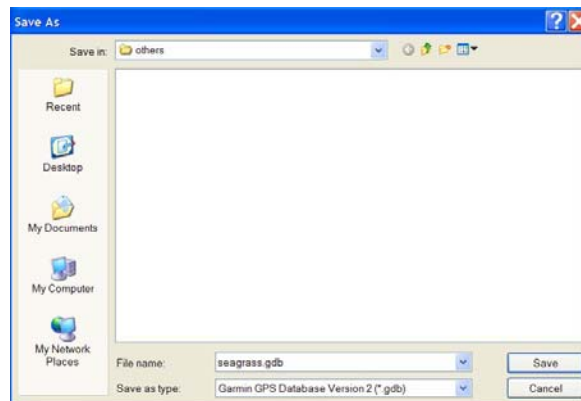
5. Click on the **Receive** button.
6. The GPS data will begin to download to the MapSource software. Click OK after data is successfully opened.

Saving GPS data in MapSource

1. To Check the GPS data that has just been downloaded, click on the “Waypoints” tab (the full list of all waypoints are listed).
2. To view the waypoints on the MapSource map area, right click on any of the waypoints (a context menu will appear) and select “**Show Selected Waypoint On Map**”. The waypoint will be shown on the map



3. You must always save the GPS data that has been downloaded to MapSource. To save the GPS data in MapSource, Click on “**File**” menu, then select “**Save**”. A “Save As” window will appear. Browse to the directory where you want to save the file in and type in a file name. Click “Save” and the MapSource file is save (this file contains all the GPS data).



The advantages for saving the GPS data are:

- GPS data will not be lost when the data is save in a MapSource file (*.mps) to your PC.
- The GPS data on the GPS unit can be deleted. The GPS unit will have the maximum memory space available for any further data capture use.
- Once the GPS data is saved as a MapSource file, the data can always be uploaded back onto the GPS unit.

This image shows a full page of white paper with horizontal dashed lines, typical of primary school writing paper. The lines are evenly spaced and run across the width of the page. There are no margins, text, or other markings on the paper.



Managing seagrass resources

Threats to seagrass habitats

Destruction or loss of seagrasses have been reported from most parts of the world, often from natural causes, e.g., "wasting disease" or high energy storms. However, destruction commonly has resulted from human activities, e.g., as a consequence of eutrophication or land reclamation and changes in land use. Increases in dredge and fill, construction on the shoreline, damage associated with commercial overexploitation of coastal resources, and recreational boating activities along with anthropogenic nutrient and sediment loading have dramatically reduced seagrass distribution in some part of the world. Anthropogenic impacts on seagrass meadows continue to destroy or degrade coastal ecosystems and decrease seagrass functions and values, including their contribution to fisheries. It is possible global climate change will have a major impact. Efforts are being made toward rehabilitation of seagrass habitat in some parts of the world: transplantation, improvement of water quality, restrictions on boating activity, fishing and aquaculture, and protection of existing habitat through law and environmental policy.

Management

Seagrasses do not exist in nature as a separate ecological component from other marine plants and are often closely linked to other community types. In the tropics the associations are likely to be complex interactions with mangrove communities and coral reef systems. In temperate waters, algae beds, salt marshes, bivalve reefs, and epiphytic plant communities are closely associated with areas of seagrass. Many management actions to protect seagrasses have their genesis in the protection of wider ecological systems or are designed to protect the overall biodiversity of the marine environment.

Seagrasses are also food for several marine mammal species and turtles, some of which are listed as threatened or vulnerable to extinction in the IUCN Red List (IUCN 2000). Seagrasses are habitat for juvenile fish and crustaceans that in many parts of the world form the basis of economically valuable subsistence and/or commercial fisheries. The need to manage fisheries in a sustainable way has itself become a motivating factor for the protection of seagrasses.

Coastal management decision making is complex, and much of the information on approaches and methods exists only in policy and legal documents that are not readily available. There may also be local or regional Government authorities having control over smaller jurisdictions with other regulations and policies that may apply. Many parts of South East Asia and the Pacific Island nations have complex issues of land ownership and coastal sea rights. These are sometimes overlaid partially by arrangements put in place by colonising powers during and after World War II, leaving the nature and strength of protective arrangements open for debate.

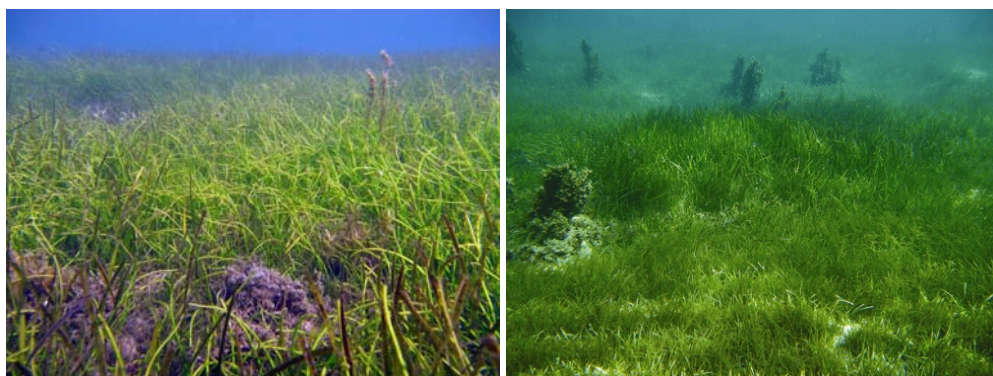
Both Australia and the United States have developed historically as Federations of States with the result that coastal issues can fall under State or Federal legislation depending on the issue or its extent. In contrast, in Europe and much of South East Asia, central Governments are more involved. Intercountry agreements in these areas such as the UNEP Strategic Action Plan for the South China Sea and the Mediterranean Countries Barcelona Convention (<http://www.unep.org/>) are required to manage marine issues that encompass more than one country.

Approaches to protecting seagrass tend to be location specific or at least nation specific (there is no international legislation directly for seagrasses as such that we know of) and depend to a large extent on the tools available in law and in the cultural approach of the community. There is, however, a global acceptance through international conventions (RAMSAR Convention; the Convention on Migratory Species of Wild Animals; and the Convention on Biodiversity) of the need for a set of standardised data/information on the location and values of seagrasses on which to base arguments for universal and more consistent seagrass protection.

We suggest that there are six precursors to successful management of coastal seagrasses:

1. Important fish habitat is known and mapped
2. Habitat monitoring is occurring
3. Adjacent catchment/watershed impacts and other threats are managed
4. Some level of public goodwill/support is present
5. Legal powers exist that are robust to challenge
6. There is effective enforcement and punishment if damage occurs

The key element is a knowledge base of the seagrass resource that needs to be protected and how stable/variable that resource is. It is also important to know if possible any areas that are of special value to the ecosystems that support coastal fisheries and inshore productivity. It is important as well that this information is readily available to decision makers in Governments in a form that can be easily understood.



Above left: *Syringodium* meadow, Cawaci, Ovalau, Jan 2007, photo by Rudi Yoshida

Above right: *Halodule uninervis* patch in *Halodule pinifolia* meadow, Tagaqe, Coral Coast, Viti Levu, Jan 2007, photo by Len McKenzie



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**We value your suggestions and any comments you may have to
improve the Seagrass-Watch program.**

Please complete the following statements in your own words

I found the Seagrass-Watch training to be

.....

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What I enjoyed most about the training was.....

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It could have been better if

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I did not realize that.....

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Now I understand that.....

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In my area the types of seagrasses and habitats include.....

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When I go back to my area, I will

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Other comments.....

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